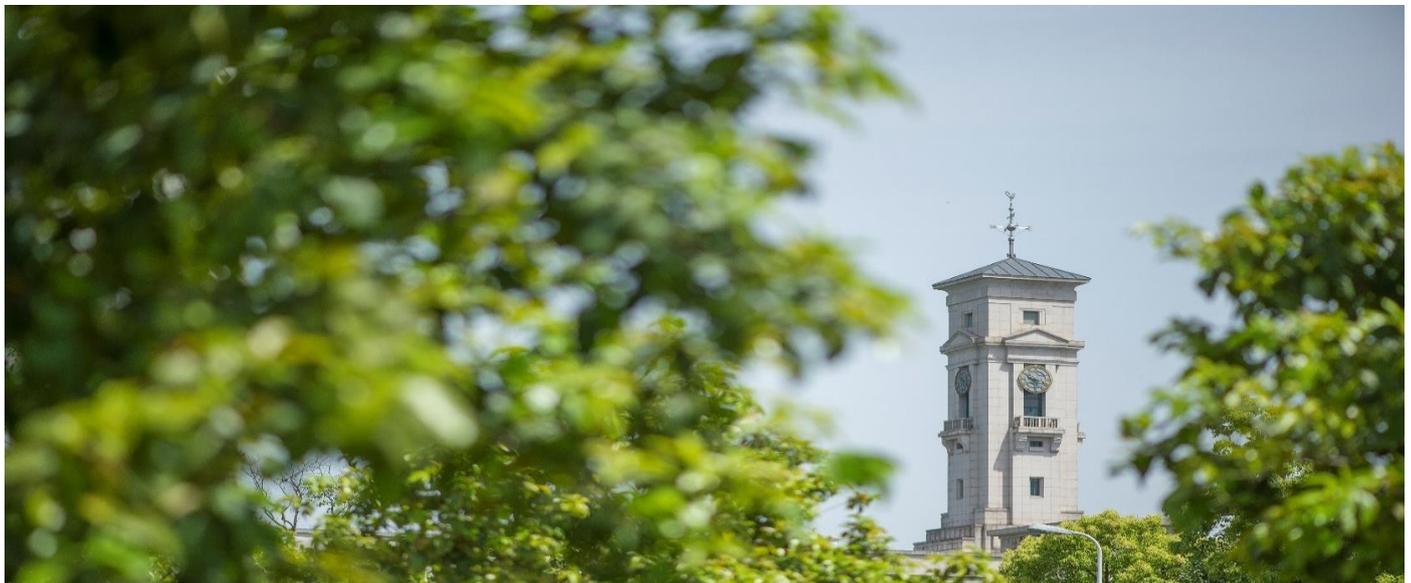


Thermophoretic collection of virus-laden (SARS-CoV-2) aerosols

Zhang A.X.^{a, b}, Wang B.J.^{c, d}, Wang C.C.^c, Lian D.Z.^e, Shi E.Y.^{a, b}, Ren F.Y.^{a, b, f}, Yan G.Y.^{a, g}



**University of
Nottingham**

UK | CHINA | MALAYSIA

University of Nottingham Ningbo China, 199 Taikang East Road, Ningbo, 315100, China

First published 2021

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Thermophoretic collection of virus-laden (SARS-CoV-2) aerosols

A. Xiangzhi Zhang,^{1,2} B. Jing Wang,^{3,4} C. Chengbo Wang,³ D. Zheng Lian,⁵ E. Yong Shi,^{1,2} F. Yong Ren^{a)},^{1,2,6} and G. Yuying Yan^{1,7}

¹⁾*Research Group for Fluids and Thermal Engineering, University of Nottingham Ningbo China, Ningbo 315100, China*

²⁾*Department of Mechanical, Materials and Manufacturing Engineering, University of Nottingham Ningbo China, Ningbo 315100, China*

³⁾*Department of Electrical and Electronic Engineering, University of Nottingham Ningbo China, Ningbo 315100, China*

⁴⁾*Key Laboratory of More Electric Aircraft Technology of Zhejiang Province, University of Nottingham Ningbo China, Ningbo 315100, China*

⁵⁾*HiFiBio (Hangzhou) Co., Ltd, Hangzhou 311215, China*

⁶⁾*Key Laboratory of Carbonaceous Wastes Processing and Process Intensification Research of Zhejiang Province, University of Nottingham Ningbo China, Ningbo 315100, China*

⁷⁾*Research Group for Fluids and Thermal Engineering, University of Nottingham, Nottingham NG7 2RD, UK*

(Dated: 24 March 2021)

Detecting the existence of SARS-CoV-2 in the indoor atmosphere is a practical solution to track the prevalence and prevent the spread of the virus. In this work, a thermophoretic approach is presented to collect the novel coronavirus-laden aerosols from the air and accumulate to high concentrations adequate for the sensitivity of viral RNA detection. Among the factors, the density and particle size have negligible effects on the particle trajectory, while the vertical coordinates of particles increase with the rise of heating source temperature. When the heating temperature is higher than 355 K, all of the particles exit the channel from one outlet, thus the collecting and accumulating of virus-laden aerosols can be realized. This study provides a potential approach to accelerate the detection of SARS-CoV-2 and avoid false negative in the following RNA test.

^{a)} Author to whom correspondence should be addressed. yong.ren@nottingham.edu.cn.

I. INTRODUCTION

The outbreak of Coronavirus disease 2019 (COVID-19) pandemic has caused a dramatic impact on healthcare services and economies in the affected countries^{1,2}. The underlying pathogen has been confirmed to be a novel coronavirus, which was named as severe acute respiratory syndrome coronavirus (SARS-CoV-2) by the International Committee on Taxonomy of Viruses³. As reported, the aerosolized virus particles carried by humans are caused by coughing/sneezing, even normal breathing or speech of an infected person⁴. The disease is asseverated to be transmitted by multiple pathways, including direct (deposited on persons) or indirect (deposited on objects) contact and airborne transmission⁵. Having analyzed the prevalence trends in China, Italy, and the United States from January 23 to May 9, 2020, Zhang et al.⁶ illustrated that airborne transmission is the dominant route to spread the disease. Respiratory particles are the media of airborne transmission, which are commonly distinguished to be droplets or aerosols based on their aerodynamic diameter⁷. According to the CDC of the US, the particles of more than $5 \mu m$ are categorized as droplets and those less than $5 \mu m$ as aerosols or droplet nuclei⁸. A study conducted by Papinei et al.⁹ showed that 80% to 90% of particles generated by human expiratory activities were aerosols, most of which were generated during coughing and the least was from nasal breathing. Up to now, the transmission mechanisms of aerosols within confined spaces are still complex and remains to be studied, especially for the indoor environment¹⁰. Highly dispersed in aerosols, the virus can stay viable and infectious for several hours¹¹. Under the condition of long exposure to high concentrations of aerosols, inhaled aerosols containing virus can deposit directly along the human respiratory tract, which causes infection in the alveolar tissues of the lower respiratory tract¹².

An aerosol is defined as a suspension system of solid or liquid particles in the air or another gas⁷. Airborne transmission can be achieved via aerosols carrying viruses, eg., the influenza A H1N1¹³, severe acute respiratory syndrome (SARS)¹⁴, and middle east respiratory syndrome (MERS)¹⁵. The indoor aerosols are most broadly defined as ultrafine ($< 0.1 \mu m$), fine ($0.1 \sim 2.5 \mu m$), or coarse ($> 2.5 \mu m$)¹⁶. Driven by Brownian motion, some of the virus-laden aerosols diffuse towards lateral directions which results in long-distance nosocomial transmission in the confined space. The transmitting payloads and environmental tolerance of SARS-CoV-2 virus of the indoor environment depend on factors including the specific phenotype available, the composition of the aerosols, and the physical characteristics of the surrounding environment¹³. Liu et al.¹⁷ investigated the generation of airborne SARS-CoV-2 and the aerosol deposition at 30

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sites in 2 designated hospitals in Wuhan, and found the concentrations of airborne SARS-CoV-2 (ranging from 16 to 42 copies per m^3) in the protective-apparel removal rooms were among the upper range.

Keeping social distancing is expected to be effective to prevent infection via bioaerosol contact¹⁸. Besides, precautions against airborne transmission in indoor scenarios should be taken including increasing ventilation rate, using natural ventilation, avoid air recirculation, avoiding staying in another person's direct airflow, and minimizing the number of people sharing the same environment¹⁹. Except for precautions, reliable diagnosis is important for epidemic prevention and control of the virus. At present, several molecular assays that detected the COVID-19 have been developed and recognized by the WHO²⁰. Among them, the reverse transcription polymerase chain reaction (RT-PCR) is commonly employed to detect the viruses and the sensitivity to detect the RNA-dependent RNA polymerase (RdRp) sequence is about 3.7 RNA copies²¹. However, the RT-PCR assays for virus detection has reported cases of false-negative results since its amplification of spurious nucleic acid contamination, and it is difficult to directly detect the viruses travelling in the air since sampling and detecting of the presence of SARS-CoV-19 is time-consuming⁵. The objective of this study is to present a novel microfluidic method to collect virus-laden aerosols from the indoor air, which enables improved sensitivity compared with existing viral detecting methods.

II. PROBLEM DESCRIPTION AND CONFIGURATION

Due to a study conducted in Singapore²², high viral RNA contained aerosols ($1.84 \times 10^3 \sim 3.38 \times 10^3$ RNA copies per m^3 air) were detected in airborne infection isolation rooms, and surface contamination were also detected in rooms with virus contained aerosols. The minimum size of SARS-CoV-2 is about 60 nm , but the combined size of virus-laden aerosol can be larger than 100 nm when attached to a larger carrier aerosol². Aerosols suspended in air collide and merge to become larger, while the shear force breaks them up. Due to the effects of coalescence and breakup, the equilibrium size is approximately 80 nm in heavy traffic area, while it can be below 100 nm in water vapour under high humidity.

In this study, a 2D model of the device is considered to investigate the performance of thermophoretic separation and collection of virus-laden aerosols. The main component of the device is a channel, the bottom wall of which is connected with a heating source. Driven by the ther-

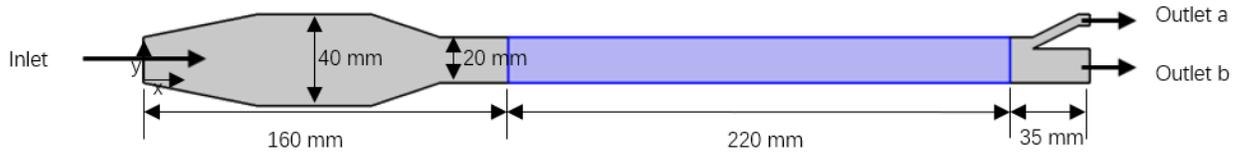


FIG. 1. Structure of device. The length of the channel is 415 mm, including an inlet region of 160 mm, a heated region of 220 mm, and an outlet region of 35 mm.

mophoretic force, the particles will move upward and exit the channel through the upper outlet. The length of the channel is 415 mm, including an inlet region of 160 mm, a thermophoresis region of 220 mm, and an outlet region of 35 mm. The geometric structure of the device is shown in Fig. 1. In the evaluation of RNA stability of SARS-CoV-2 under thermal treatment, the virus was inactivated after being heated at 60 °C for more than 15 minutes²³, which allows the thermophoretic sorting of virus-laden aerosols during a short time interval.

III. MATHEMATICAL MODEL

A. Thermophoretic force

Thermophoresis is an important transport mechanism of small particles in a non-isothermal carrier fluid driven by the temperature gradient. Particles suspended in a non-isothermal mixture subject to a forces pushing them in the direction of the temperature drop. The driving mechanism behind this force is the collision of gas molecules on the particles surface. Collisions are more likely to occur on the hotter side of the particle where the average molecular velocity of the gas is greater. This results in a net force towards colder regions of the gas. Particle with different physical properties exhibit different responses to the force, which realises sorting and separation. In gas media, the thermophoretic force acting on a suspended particle depends on the flow regime characterized by the Knudsen number $Kn = \lambda/L$, where λ is the mean molecular free path and L is the characteristic length of the particle. For small particle or large molecular mean free path when $Kn \gg 1$, the effect of the particle motion on the distribution of the fluid molecular velocities can be virtually neglected. Whereas, solving the Boltzmann equations in continuum ($Kn \ll 1$) and transition ($Kn \approx 1$) regimes where the velocity distribution of molecules is greatly affected by the

movement of particles presents considerable complexities, since rarefied gas dynamics remains to be resolved by kinetic theory. Epstein²⁴ derived an equation for the thermophoretic force exerted on spherical particles in gases based on a continuum analysis:

$$\mathbf{F}_{\text{tp}} = -\frac{4.5\pi d_p \mu^2}{\rho} \frac{1}{2 + \frac{k_p}{k}} \frac{\nabla T}{T} \quad (1)$$

where d_p is the diameter of particle, μ is the coefficient of shear viscosity, ρ is the mass density, $\frac{k_p}{k}$ is the thermal conductivity ratio of particle and fluid, T is the absolute temperature. Since boundary conditions appropriate for the slip-flow regime have not been used and the continuum energy equation has been solved regardless of the convective terms, serious disagreement between Eq. (1) and experimental results²⁵. Brock²⁶ conducted a hydrodynamic analysis for small $Kn \ll 1$ in near continuum regime and developed a general equation with introducing matching coefficients associated with the temperature jump and velocity slip. Talbot et al.²⁷ established an equation to describe the thermophoretic force for the entire range of Kn :

$$\mathbf{F}_{\text{tp}} = -\frac{6\pi\mu^2 d_p C_s \left(\frac{k}{k_p} + C_t \frac{2\lambda}{d_p} \right) \frac{\nabla T}{T}}{\rho \left(1 + 6C_m \frac{\lambda}{d_p} \right) \left(1 + 2\frac{k}{k_p} + 4C_t \frac{\lambda}{d_p} \right)} \quad (2)$$

where the matching parameters $C_s = 1.17$, $C_m = 1.14$, $C_t = 2.18$. According to the Stokes expression, the thermophoretic velocity is obtained as

$$\mathbf{u}_{\text{tp}} = -\frac{2\eta C_s \left(\frac{k}{k_p} + C_t \frac{2\lambda}{d_p} \right) \frac{\nabla T}{T}}{\rho \left(1 + 6C_m \frac{\lambda}{d_p} \right) \left(1 + 2\frac{k}{k_p} + 4C_t \frac{\lambda}{d_p} \right)} \quad (3)$$

where η is the dynamic viscosity of the gas.

B. Governing equations and boundary conditions

This model is composed of flow field, temperature field, and particle tracing. The flow field is characterized by Navier-Stokes equations:

$$\begin{cases} \nabla \cdot (\rho \mathbf{u}) = 0 \\ \rho(\mathbf{u} \cdot \nabla) \mathbf{u} = \nabla \cdot [-p\mathbf{I} + \mu(\nabla \mathbf{u} + (\nabla \mathbf{u})^\top) - \frac{2}{3}\mu(\nabla \cdot \mathbf{u})\mathbf{I}] + \mathbf{F} \end{cases} \quad (4)$$

where ρ is the density, \mathbf{u} is the velocity, p is the pressure, \mathbf{I} is the identity matrix, μ is the viscosity, \mathbf{F} is the force term.

Sample title

Heat transfer in fluids is characterized by:

$$\rho c_p \frac{\partial T}{\partial t} + \rho c_p \mathbf{u} \cdot \nabla T + \nabla \cdot (-k \nabla T) = Q \quad (5)$$

where c_p is the heat capacity, T is the temperature, Q is the heat flux term.

The trajectory of the particle in fluid flow is characterized by:

$$\frac{d(m_p \mathbf{u})}{dt} = \mathbf{F}_p \quad (6)$$

where \mathbf{F}_p is the force of particles. Particles in aerosols are often subjected to Brownian motion, gravity, electrostatic forces, thermal gradients, electromagnetic radiation, turbulent diffusion, and inertial forces²⁸. Buongiorno et al.²⁹ investigated the relative effects of inertia, Brownian diffusion, thermophoresis, diffusiophoresis, Magnus effect, fluid drainage, and gravity that may cause a relative motion of particles in the main fluid. For particles at nanoscale, only thermophoresis and Brownian diffusion can cause slip. For micro-sized particles, gravity (weight and buoyancy) should be considered, while Brownian diffusion is less important. In continuum mechanics, the Froude number is a dimensionless number defined as the ratio of the flow inertia to the external field, expressed as

$$Fr = \frac{u}{\sqrt{gL}} \quad (7)$$

where u is the magnitude of local flow velocity, g is the magnitude of the gravity field, L is a characteristic length. In this study, the Froude number $Fr \ll 1$ and the flow in channel is a subcritical flow³⁰, thus the source term in Eq. (6) can be expressed as

$$\mathbf{F}_p = \mathbf{F}_b + \mathbf{F}_g + \mathbf{F}_d + \mathbf{F}_{tp} \quad (8)$$

where, \mathbf{F}_b , \mathbf{F}_g , \mathbf{F}_d , \mathbf{F}_{tp} represents the Brownian force, gravity force, drag force, and thermophoretic force, respectively.

Brownian motion is the random, uncontrolled movement of particles in a fluid as they constantly collide with other molecules, which leads to spreading of particles from regions of high particle density to low density³¹. The total force on the particles undergo Brownian motion is expressed by a Brownian force term \mathbf{F}_b

$$\mathbf{F}_b = \zeta \sqrt{\frac{12\pi k_B \mu T r_p}{\Delta t}} \quad (9)$$

where ζ is a normally distributed random number with a mean of zero, $k_B = 1.380649 \times 10^{-23} \text{ J/K}$ is the Boltzmann constant, μ is the fluid dynamic viscosity, T is the absolute fluid temperature, r_p

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is the particle radius, Δt is the time step taken by the solver. Gravity force is expressed by

$$\mathbf{F}_g = m_p \mathbf{g} \frac{\rho_p - \rho}{\rho_p} \quad (10)$$

where m_p is the mass of particle, \mathbf{g} is the gravity vector, ρ_p is the density of particle. \mathbf{F}_d represents the drag force expressed as

$$\mathbf{F}_d = \frac{18\mu}{\rho_p d_p^2} m_p \mathbf{u}_r \quad (11)$$

where ρ_p , d_p represents the density or diameter of the particle, respectively; $\mathbf{u}_r = \mathbf{u} - \mathbf{u}_p$ is the relative velocity. And the thermophoretic force exerted on aerosol particles is expressed as Eq. (2).

Air and aerosols are injected into the domain, carried by an air flow of 2000 Q_{scm} (Standard Cubic Centimeters per minute). The standard molar volume $V_m = 0.0224136 \text{ m}^3/\text{mol}$, the mean molar mass $M_n = 0.002 \text{ kg/mol}$. The diameter of the inlet is 20mm, with temperature 293.15K. The boundary condition of the outlet has a relative pressure $p = 0$. After entering the thermophoresis section, the particles will be heated by susceptor (with temperature T_{sus}), and then migrate to the upper wall due to the impact of thermophoresis. The heat convection with the environment ($T_e = 293.15 \text{ K}$) is through the other walls (the thickness $d = 5 \text{ mm}$, the heat transfer coefficient $h = 10 \text{ W/m}^2\text{K}$).

IV. RESULTS AND DISCUSSION

A. Numerical validation

The numerical simulation is conducted on a desktop PC with Intel(R) Core(TM) i7-6700 CPU and 32 GB physical RAM. Table I shows the number of elements, minimum element quality, and the number of degree of freedom (DOF) with varying grid densities. According to this table, the minimum element quality is greater than 0.1 when the number of domain elements is greater than 1286. Therefore, numerical simulation is conducted on the mesh with 884 boundary elements and 15718 domain elements.

The validation of the model presented in this study is investigated by comparing with the results of Eslamian et al.³² for precise particle thermophoretic separation and manipulation in microchannels. In their study, a two-dimensional microchannel designs with a width of 500 μm and length of 8 mm with one inlet and two outputs are considered. Fig. 2 shows that the agreement of results is generally well.

TABLE I. Elements numbers, minimum element quality and degree of freedom (DOF) with varying grid densities.

Boundary elements	Domain elements	Minimum element quality	Degree of freedom(DOF)
211	1286	0.1849	3802
305	2134	0.1633	6083
382	3311	0.2029	8930
559	6187	0.2021	15808
707	9415	0.1877	23210
884	15718	0.2052	36927
1820	41019	0.2144	93571
3484	104354	0.2117	230995
3516	162422	0.2074	347199

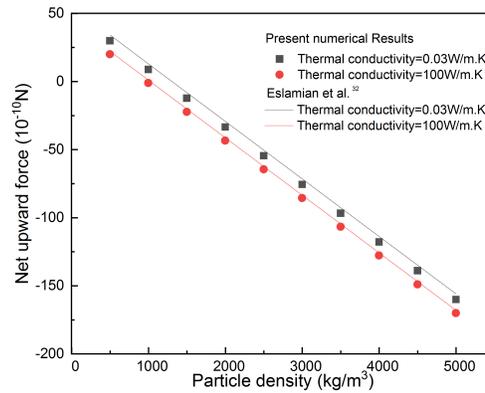


FIG. 2. Comparison of net upward force variations along particle density.

B. Fields and thermophoresis performance

In this study, the numerical simulations are conducted to investigate the particle trajectories of aerosols in a laminar flow of carrier fluid in a channel shown in Fig. 1. The flow of air in the channel with aerosol particles injected is shown in Fig. 3 and Fig. 4. According to Fig. 3, the global pressure drop is $3.28 \times 10^{-4} Pa$. The maximum Reynolds number is approximately 0.05, the flow regime in this channel is creeping flow. Shown in Fig. 4, the average radial velocity

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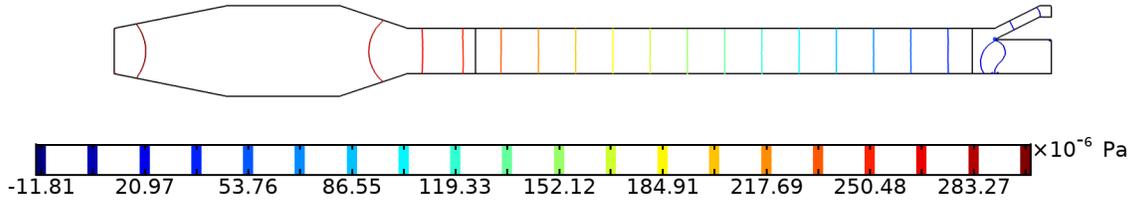


FIG. 3. Pressure distribution in the channel with air flow of $2000 Q_{sccm}/m$ at the inlet.

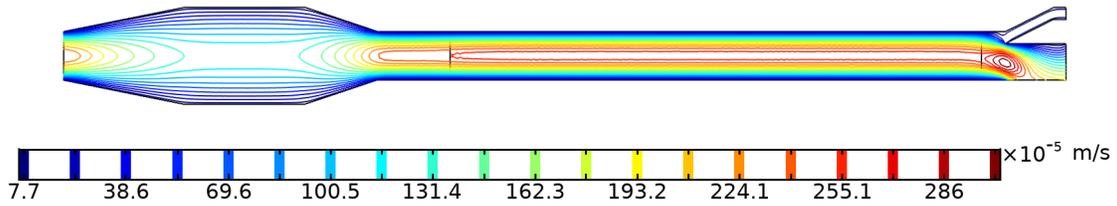


FIG. 4. Distribution of velocity magnitude with air flow of $2000 Q_{sccm}/m$ at the inlet.

$u_a = 0.0014 m/s$. The flowrate at the upper outlet: $v_a = 4.9412 \times 10^{-7} m^2/s$; while the flow rate at the lower outlet: $v_b = 1.6076 \times 10^{-5} m^2/s$. The ratio of outlet flowrate $v_a/v_b = 0.02887$, the particle concentration is raised at approximately 35 times. The temperature distribution when the bottom wall is connected with a heating source of $320 K$ is shown in Fig. 5.

The real-time trajectories aerosol particles are shown in Fig. 6 and Fig. 7. Due to the thermal gradient, aerosols are driven by thermophoretic force to move upward, and then exit the channel through the upper outlet. On the horizontal direction, the particles are carried by the inertia force and move right with the carrier fluid. On the vertical direction, the particles are driven by buoyancy and thermophoretic forces to move upward, whereas the gravity acts downward. Gravity and buoyancy forces are proportional to the particle diameter cubed while the thermophoretic force is linearly proportional to the particle diameter. Since the density of aerosol particles are close

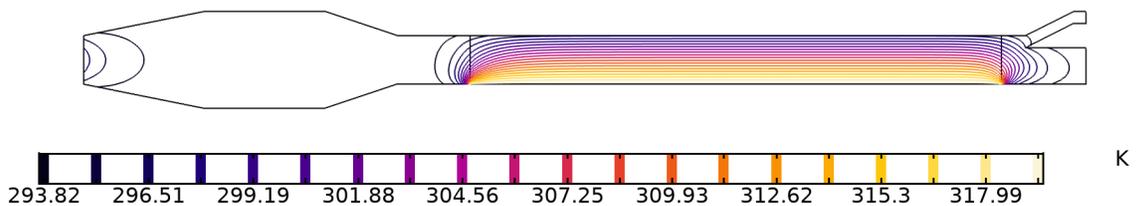


FIG. 5. Distribution of temperature when $T_{susc} = 320K$.

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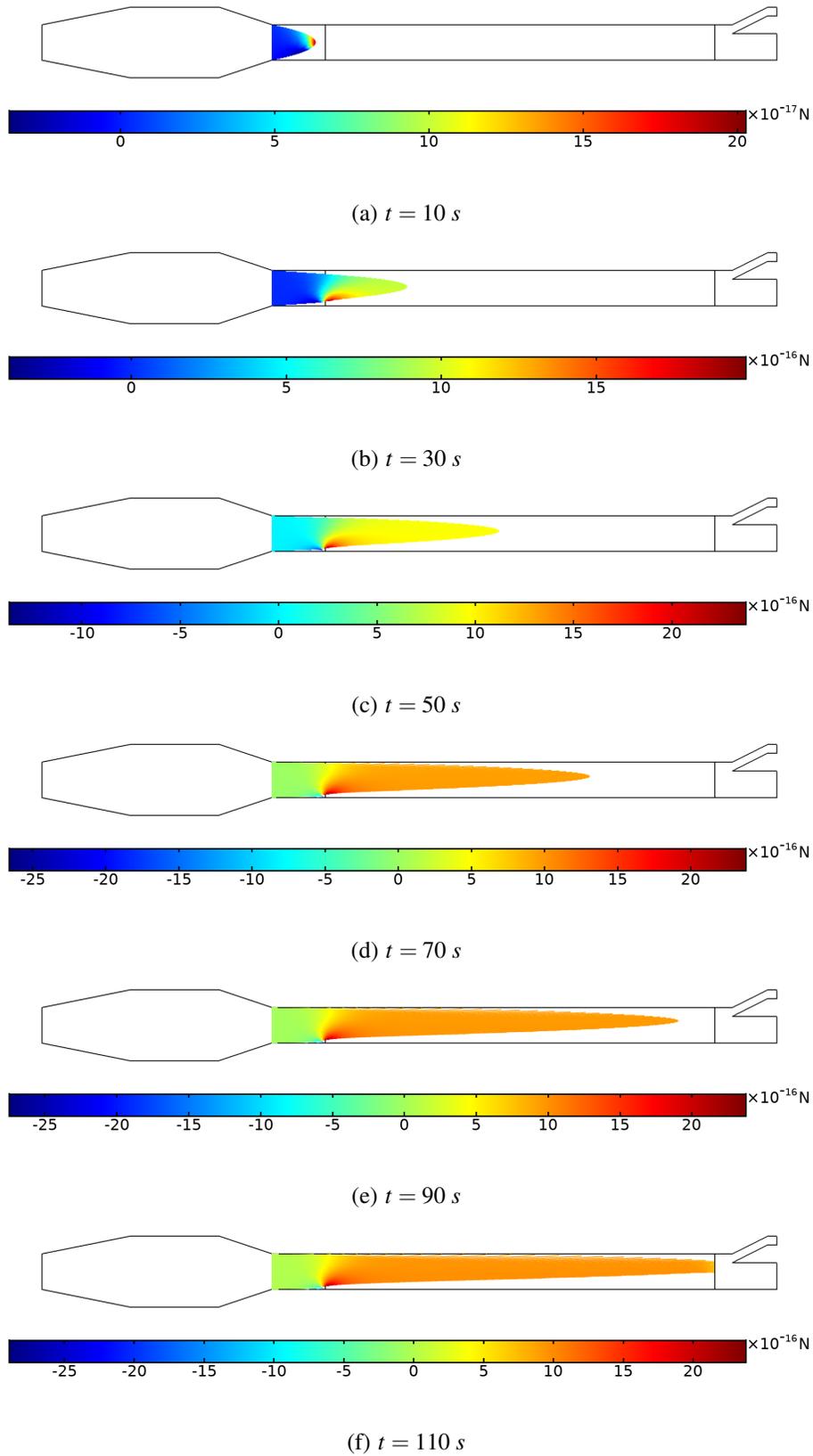


FIG. 6. Real-time positions of aerosol particles when $T_{susc} = 320 \text{ K}$, the colour legend represents the vertical component of thermophoretic force.

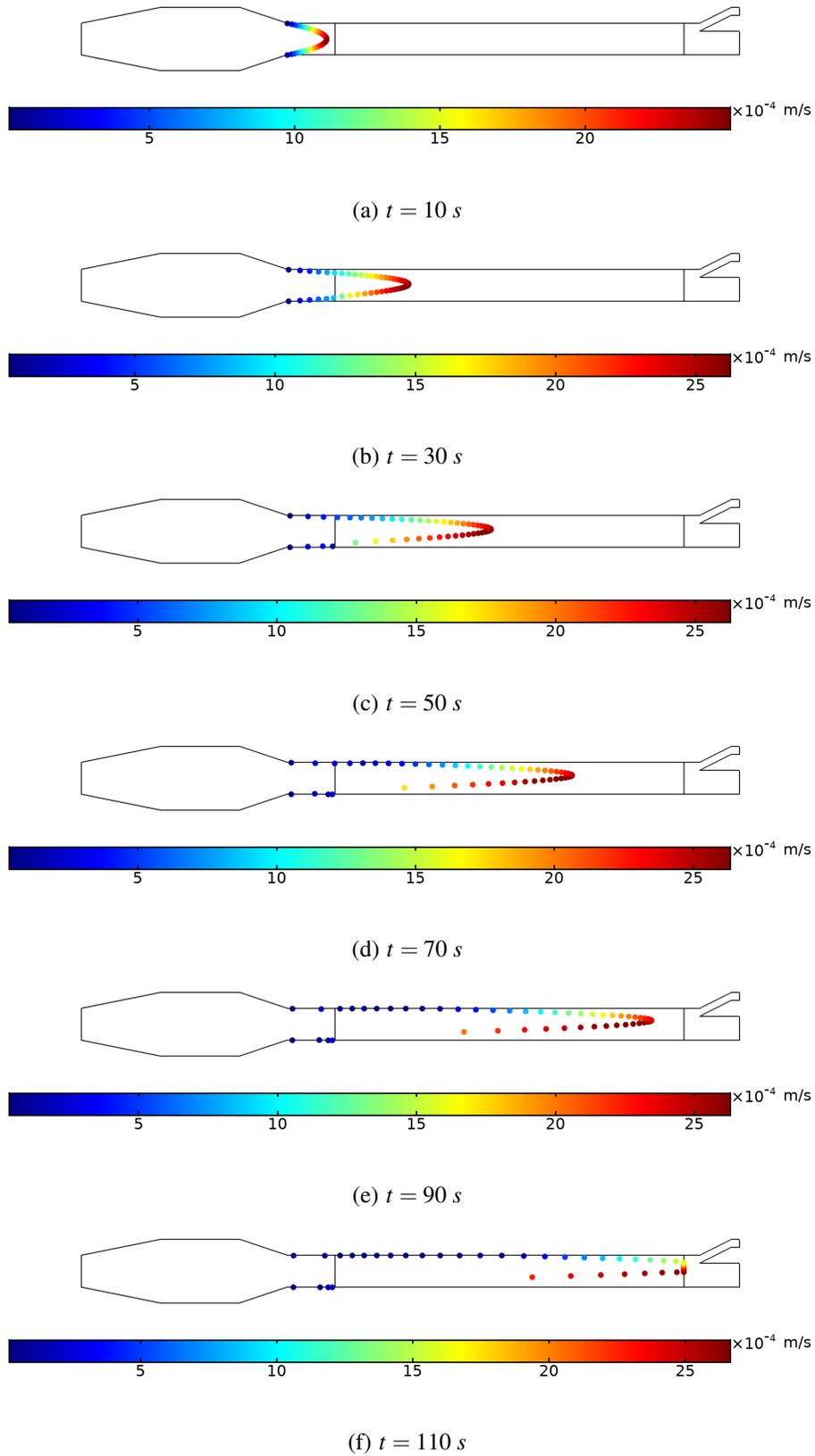


FIG. 7. Real-time positions of aerosol particles when $T_{susc} = 320\text{ K}$, the colour legend represents the vertical velocity.

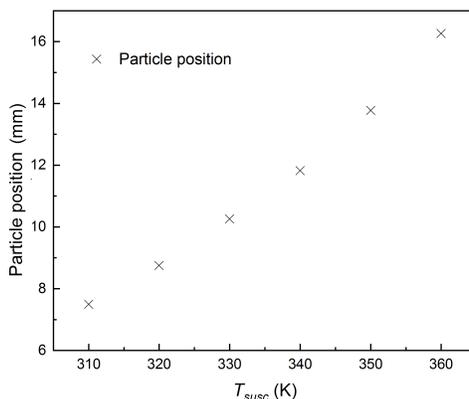


FIG. 8. The minimum vertical coordinates of particles at the outlet with heating source temperatures.

to the carrier fluid, the buoyancy can be almost balanced by the gravity. Therefore, the trajectories of particles with varying sizes showed a tiny divergence on the vertical direction, while the density and particle size have negligible effect on the particle trajectory. According to Eq. (6), thermophoretic force is also determined by the local temperature and its gradient. The minimum vertical coordinates (infima) of aerosols at the outlet with varying heating source temperatures are shown in Fig. 8. According to Fig. 8, the vertical coordinates of aerosols increase with the rise of heating source temperature. When the temperature is higher than 355 K, the minimum vertical coordinate of particles is 15 mm, which means all of the particles exit the channel from the upper outlet, thus the collecting and accumulating of virus-laden aerosols are achieved. The concentration of virus-laden aerosols can be raised to 25 times after thermophoretic sorting, thus enhancing the RNA concentration, which can be beneficial to reduce the possibility of false negative in RNA test.

V. CONCLUSION

In this study, thermophoresis is adopted as a novel technique to collect and accumulate virus-laden aerosols from indoor air. Numerical simulations were conducted by coupling the transfer equations of heat and mass transfer with particle tracing for fluid flow. The availability of this device was verified and suitable operation conditions were determined based on the simulation and the parameters acquired from articles and experiments. It has been acquired that thermophoretic force is significant to drive the virus-bearing aerosols to move upward and accumulate at the

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cold side. When the heating temperature is higher than 355K, all the particles exit the channel through the upper outlet, thus achieving the collecting of virus-laden aerosols. Considering the stability of viral RNA, a suitable range (360 ~ 380 K) of heating source temperature is acquired. Under these operation conditions, the aerosols can be collected from indoor air and accumulated to a higher density which beyond the test requirements without being destroyed by heat. This technique provides a method to reduce the time interval of the RNA test of SARS-CoV-2 and avoid false negative for RNA detection, which is helpful to suppress the spread of the pandemic of Covid-19.

AUTHOR'S CONTRIBUTION

X. Zhang and Y. Ren proposed the research and designed the numerical work, X. Zhang conducted the simulation, X. Zhang and Y. Ren wrote the manuscript, J. Wang, C. Wang, Z. Lian, Y. Shi and Y. Yan contributed to the discussion of the data, and review of the manuscript.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ACKNOWLEDGMENTS

This work was financially supported by Zhejiang Provincial Natural Science Foundation of China under Grant No. LY19E060001 and LQ19F050003, Ningbo Science and Technology Bureau under Service Industry Science & Technology Programme with project code 2019F1030, as well as EU ThermaSMART project under Grant No. H2020-MSCA-RISE (778104)-Smart thermal management of high power microprocessors using phase-change (ThermaSMART). The project was also supported by Ningbo Science and Technology Bureau Technology Innovation Team (grant no. 2016B10010). The Zhejiang Provincial Department of Science and Technology is acknowledged for this research under its Provincial Key Laboratory Programme (2020E10018).

REFERENCES

- ¹W. Tan, X. Zhao, X. Ma, W. Wang, P. Niu, W. Xu, G. F. Gao, and G. Wu, “A novel coronavirus genome identified in a cluster of pneumonia cases—Wuhan, China 2019 2020,” *China CDC Weekly* **2**, 61–62 (2020).
- ²P. Zhou, X.-L. Yang, X.-G. Wang, B. Hu, L. Zhang, W. Zhang, H.-R. Si, Y. Zhu, B. Li, and C.-L. Huang, “A pneumonia outbreak associated with a new coronavirus of probable bat origin,” *nature* **579**, 270–273 (2020).
- ³C. S. G. of the International, “The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2,” *Nature Microbiology* **5**, 536 (2020).
- ⁴N. H. L. Leung, D. K. W. Chu, E. Y. C. Shiu, K.-H. Chan, J. J. McDevitt, B. J. P. Hau, H.-L. Yen, Y. Li, D. K. M. Ip, and J. S. M. Peiris, “Respiratory virus shedding in exhaled breath and efficacy of face masks,” *Nature medicine* **26**, 676–680 (2020).
- ⁵L. Morawska and J. Cao, “Airborne transmission of SARS-CoV-2: The world should face the reality,” *Environment International* , 105730 (2020).
- ⁶R. Zhang, Y. Li, A. L. Zhang, Y. Wang, and M. J. Molina, “Identifying airborne transmission as the dominant route for the spread of COVID-19,” *Proceedings of the National Academy of Sciences* (2020).
- ⁷W. C. Hinds, *Aerosol technology: properties, behavior, and measurement of airborne particles* (John Wiley & Sons, 1999).
- ⁸E. Y. C. Shiu, N. H. L. Leung, and B. J. Cowling, “Controversy around airborne versus droplet transmission of respiratory viruses: implication for infection prevention,” *Current opinion in infectious diseases* **32**, 372–379 (2019).
- ⁹R. S. Papineni and F. S. Rosenthal, “The size distribution of droplets in the exhaled breath of healthy human subjects,” *Journal of Aerosol Medicine* **10**, 105–116 (1997).
- ¹⁰T. Wang, C. Lien, S. Liu, and P. Selveraj, “Effective Heat Inactivation of SARS-CoV-2,” *medRxiv* , 2020.04.29.20085498 (2020).
- ¹¹N. Van Doremalen, T. Bushmaker, D. H. Morris, M. G. Holbrook, A. Gamble, B. N. Williamson, A. Tamin, J. L. Harcourt, N. J. Thornburg, and S. I. Gerber, “Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1,” *New England Journal of Medicine* **382**, 1564–1567 (2020).

Sample title

- ¹²R. J. Thomas, “Particle size and pathogenicity in the respiratory tract,” *Virulence* **4**, 847–858 (2013).
- ¹³M. Schuit, S. Gardner, S. Wood, K. Bower, G. Williams, D. Freeburger, and P. Dabisch, “The influence of simulated sunlight on the inactivation of influenza virus in aerosols,” *The Journal of infectious diseases* **221**, 372–378 (2020).
- ¹⁴Y. Li, X. Huang, I. Yu, T. Wong, and H. Qian, “Role of air distribution in sars transmission during the largest nosocomial outbreak in hong kong,” *Indoor air* **15**, 83–95 (2005).
- ¹⁵N. Van Doremalen, T. Bushmaker, and V. J. Munster, “Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions,” *Eurosurveillance* **18**, 20590 (2013).
- ¹⁶W. W. Nazaroff, “Indoor particle dynamics,” *Indoor air* **14**, 175–183 (2004).
- ¹⁷Y. Liu, Z. Ning, Y. Chen, M. Guo, Y. Liu, N. K. Gali, L. Sun, Y. Duan, J. Cai, and D. Westerdahl, “Aerodynamic analysis of SARS-CoV-2 in two Wuhan hospitals,” *Nature* **582**, 557–560 (2020).
- ¹⁸M. Guzman, “Bioaerosol size effect in COVID-19 transmission,” (2020).
- ¹⁹H. Qian and X. Zheng, “Ventilation control for airborne transmission of human exhaled bioaerosols in buildings,” *Journal of thoracic disease* **10**, S2295 (2018).
- ²⁰W. H. Organization, en“Laboratory testing for coronavirus disease 2019 (COVID-19) in suspected human cases: interim guidance, 2 March 2020,” Tech. Rep. (Geneva PP - Geneva).
- ²¹V. M. Corman, O. Landt, M. Kaiser, R. Molenkamp, A. Meijer, D. K. Chu, T. Bleicker, S. Brünink, J. Schneider, M. L. Schmidt, D. G. Mulders, B. L. Haagmans, B. van der Veer, S. van den Brink, L. Wijsman, G. Goderski, J.-L. Romette, J. Ellis, M. Zambon, M. Peiris, H. Goossens, C. Reusken, M. P. Koopmans, and C. Drosten, eng“Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR,” *Euro surveillance : bulletin Europeen sur les maladies transmissibles = European communicable disease bulletin* **25**, 2000045 (2020).
- ²²P. Y. Chia, K. K. Coleman, Y. K. Tan, S. W. X. Ong, M. Gum, S. K. Lau, X. F. Lim, A. S. Lim, S. Sutjipto, P. H. Lee, T. T. Son, B. E. Young, D. K. Milton, G. C. Gray, S. Schuster, T. Barkham, P. P. De, S. Vasoo, M. Chan, B. S. P. Ang, B. H. Tan, Y.-S. Leo, O.-T. Ng, M. S. Y. Wong, K. Marimuthu, D. C. Lye, P. L. Lim, C. C. Lee, L. M. Ling, L. Lee, T. H. Lee, C. S. Wong, S. Sadarangani, R. J. Lin, D. H. L. Ng, M. Sadasiv, T. W. Yeo, C. Y. Choy, G. S. E. Tan, F. Dimatatac, I. F. Santos, C. J. Go, Y. K. Chan, J. Y. Tay, J. Y.-L. Tan, N. Pandit, B. C. H. Ho, S. Mendis, Y. Y. C. Chen, M. Y. Abdad, D. Moses, and f. t. S. . N. C. O. R. Team, “Detection of air and surface contamination by SARS-CoV-2 in hospital rooms of infected patients,” *Nature*

Sample title

Communications **11**, 2800 (2020).

- ²³J. Wang and G. Du, “COVID-19 may transmit through aerosol,” *Irish Journal of Medical Science* (1971-), 1–2 (2020).
- ²⁴P. Epstein, “Zeitschrift für physik a,” *Zur Theorie des Radiometers* **54**, 537–563 (1929).
- ²⁵C. Schadt and R. Cadle, “Thermal forces on aerosol particles1,” *The Journal of Physical Chemistry* **65**, 1689–1694 (1961).
- ²⁶J. R. Brock, “On the theory of thermal forces acting on aerosol particles,” *Journal of Colloid Science* **17**, 768–780 (1962).
- ²⁷L. Talbot, R. Cheng, R. Schefer, and D. Willis, “Thermophoresis of particles in a heated boundary layer. 101,” (1980).
- ²⁸P. A. Baron and K. Willeke, “Aerosol Measurement: Principles, Techniques and Applications 2001,”.
- ²⁹J. Buongiorno, “Convective transport in nanofluids,” (2006).
- ³⁰K. W. Nicholson, “Physical Aspects of Bioaerosol,” *Bioaerosols handbook* , 27 (1995).
- ³¹J. G. Mitchell and K. Kogure, “Bacterial motility: links to the environment and a driving force for microbial physics,” *FEMS microbiology ecology* **55**, 3–16 (2006).
- ³²M. Eslamian and M. Z. Saghir, “Novel thermophoretic particle separators: numerical analysis and simulation,” *Applied thermal engineering* **59**, 527–534 (2013).